

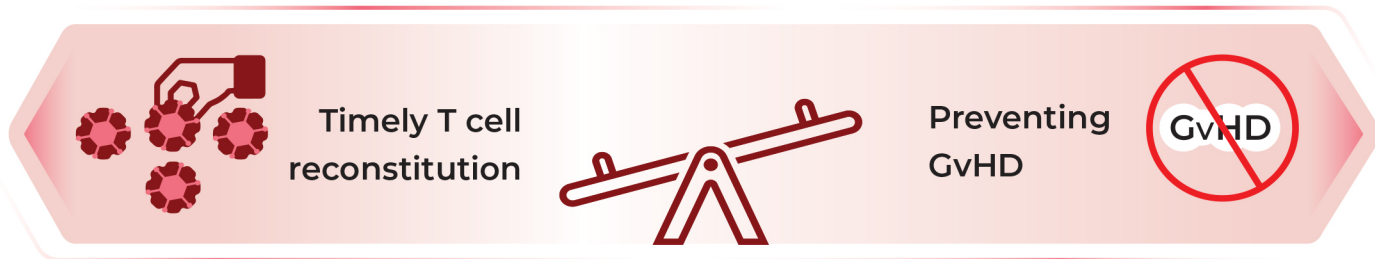


The Hemisphere Tribune

Updates and Insights on r-ATG in BMT

Individualizing rabbit anti-thymocyte globulin (r-ATG) exposure in every patient may be the key to overcoming the drawbacks of current weight-based r-ATG dosing strategies

Dose of r-ATG is pivotal in maintaining a balance.¹

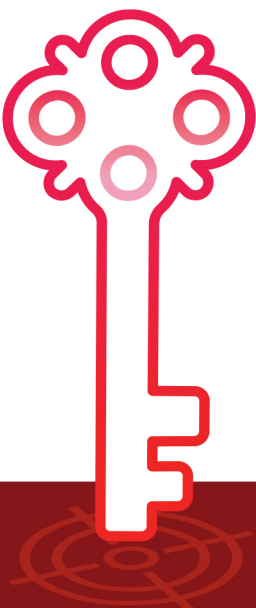


Challenges related to the strategy:²⁻⁵

Weight-based r-ATG dosing strategies in HSCT patients¹⁻⁴



- ▶ Poor T cell reconstitution
- ▶ Reduced graft-versus-leukemia effect and increased relapse and non-relapse mortality



The current weight-based dosing is empiric at its best!



Hence, there is a need for individualized r-ATG dosing for reducing exposure to r-ATG.

As per USFDA and EMA guidelines:¹

Population pharmacokinetics modeling

is required to determine an optimum individual r-ATG exposure.

Individualized optimum r-ATG exposure as determined by the pharmacokinetic modeling may hold the key to optimal patient outcomes with r-ATG dosing.^{6,7}

BMT: Bone marrow transplantation; EMA: European Medicines Agency; GvHD: Graft-versus-host disease; HSCT: Hematopoietic stem cell transplantation; USFDA: United States Food and Drug Administration.



Association between r-ATG exposure and CD4⁺ immune reconstitution⁸

AUC of r-ATG



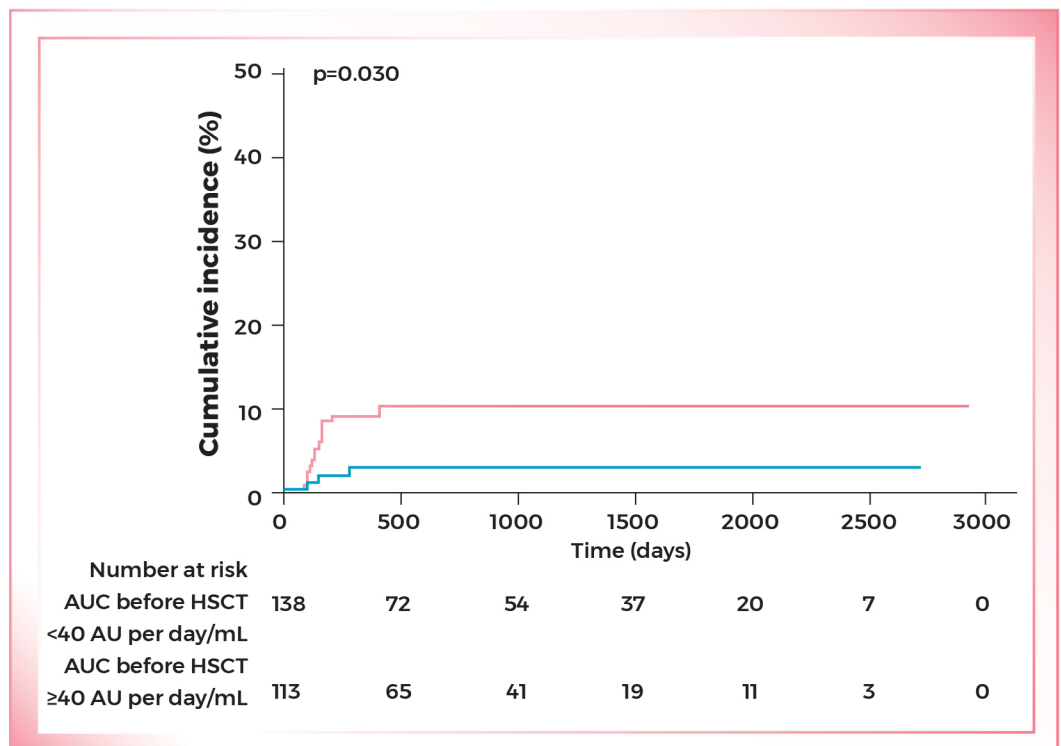
Pharmacokinetic modeling studies and clinical trials demonstrate a strong relationship between the r-ATG exposure and T cell reconstitution⁸



Immune reconstitution post-HSCT

Study design and population	Outcome 1	Outcome 2	Conclusion
Retrospective analysis of pediatric aHSCT patients by Admiraal <i>et al.</i> ⁸ (N=251)	Reduced chance of successful immune reconstitution with increasing r-ATG AUC after aHSCT (odds ratio: 0.991; p<0.0001)	r-ATG AUC threshold minimum of 40 AU per day/mL significantly reduced aGvHD and cGvHD	Effective immune reconstitution increases OS and decreases NRM and relapse mortality

Cumulative incidence curves for graft failure according to AUC before transplantation⁸

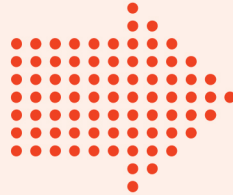


aGvHD: Acute graft-versus-host disease; aHSCT: Allogeneic hematopoietic stem cell transplantation; AU: Arbitrary unit; AUC: Area under the curve; CD4⁺: Cluster of differentiation 4⁺; cGvHD: Chronic graft-versus-host disease; HSCT: Hematopoietic stem cell transplantation; NRM: Non-relapse mortality; OS: Overall survival; r-ATG: Rabbit anti-thymocyte globulin.

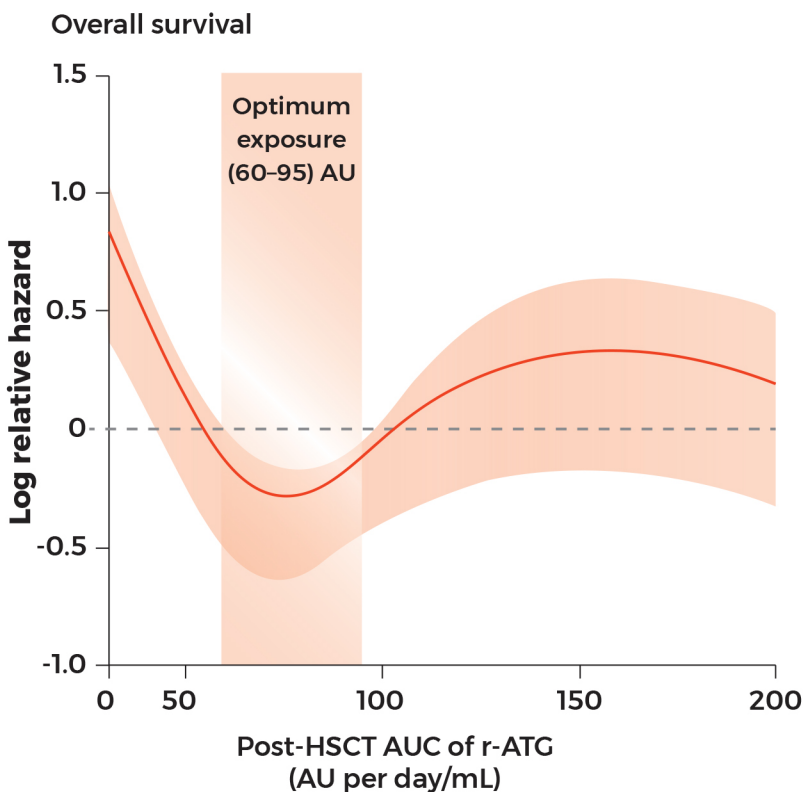
Association between r-ATG exposure and the 5-year overall survival outcome¹



Retrospective analysis of 146 ALL, AML, and MDS patients¹



Optimum r-ATG exposure post-transplantation= 60-95 AU per day/mL



Optimum r-ATG exposure after HSCT for favorable overall survival¹

Other outcomes

Relapse mortality

Non-relapse mortality

r-ATG exposure after aHSCT—Best predictor of 5-year survival

The model revealed that ALC-modeled dosing is better than weight-based dosing for optimal target exposure of r-ATG, especially in patients >50 kg.¹



The model-suggested cumulative intravenous r-ATG dosage for optimal r-ATG exposure can be calculated using:¹

$$\text{Cumulative dose} = 400 + 350 \times \text{lymphocyte count (in } 10^9/\text{L)}$$

aHSCT: Allogeneic hematopoietic stem cell transplantation; ALC: Absolute lymphocyte count; ALL: Acute lymphoid leukemia; AML: Acute myeloid leukemia; AU: Arbitrary unit; AUC: Area under the curve; BMT: Bone marrow transplantation; HSCT: Hematopoietic stem cell transplantation; MDS: Myelodysplastic syndrome; r-ATG: Rabbit anti-thymocyte globulin.

Prospective clinical evidences on individualization of r-ATG dose and aHSCT outcomes

PARACHUTE study⁹

aHSCT pediatric patients were treated with **individualized dosing regimen** based on body weight and ALC at the time of r-ATG initiation.

Primary endpoint: T cell reconstitution $>0.05 \times 10^9$ CD4⁺ T cells/L twice within 100 days [± 3] after transplantation

Outcome⁹

ALC-based dosing regimen led to optimum r-ATG exposure in 97% vs. 30%–53% of the patients via weight-based dosing regimen.⁹

80%

of the subjects met the primary endpoint within 100 days with individualized r-ATG dosing.

20% more than weight-based dosing from historical cohort

85%

Other similar studies using **weight- and ALC-based r-ATG nomogram dosing** have achieved their primary endpoint.^{10,11}

88%

Patients with inborn error of metabolism¹⁰

T cell reconstitution in pediatric aHSCT

Patients¹¹

Single-centered prospective study¹²

Optimal active r-ATG exposure associated with the minimum risk of virus reactivation and grade II–IV aGvHD¹²

Adult patients with hematological malignancies undergoing haplo-PBSCT¹²

Wang *et al.* identified that **optimal active r-ATG 100–148.5 UE/mL/day is associated with the minimum risk of virus reactivation and grade II–IV aGvHD.¹²**

Conclusion: Emerging clinical outcomes from attempts at individualizing r-ATG dose based on ALC±weight-guided r-ATG exposure are quite encouraging and lend credence to further evaluation attempts in the form of large-scale RCTs.

aGvHD: Acute graft versus host disease; aHSCT: Allogeneic hematopoietic stem cell transplantation; CD4+: Cluster of differentiation 4+; PBSCT: Peripheral blood stem cell transplantation; r-ATG: Rabbit anti-thymocyte globulin; RCTs: Randomized clinical trials.

References: 1. Lindemans CA, Chiesa R, Amrolija PJ, *et al.* Impact of thymoglobulin prior to pediatric unrelated umbilical cord blood transplantation on immune reconstitution and clinical outcome. *Blood*. 2014;123:126–132. 2. Kröger N, Solano C, Wolschke C, *et al.* Antilymphocyte globulin for prevention of chronic graft-versus-host disease. *N Engl J Med*. 2016;374:43–53. 3. Bartelink IH, Belitser SV, Knibbe CAJ, *et al.* Immune reconstitution kinetics as an early predictor for mortality using various hematopoietic stem cell sources in children. *Biol Blood Marrow Transplant*. 2013;19:305–313. 4. Bosch M, Dhadda M, Hoegh-Petersen M, *et al.* Immune reconstitution after anti-thymocyte globulin-conditioned hematopoietic cell transplantation. *Cytotherapy*. 2012;14:1258–1275. 5. Admiraal R, Nierkens S, de Witte MA, *et al.* Association between anti-thymocyte globulin exposure and survival outcomes in adult unrelated haemopoietic cell transplantation: A multicentre, retrospective, pharmacodynamic cohort analysis. *Lancet Haematol*. 2017;4(4):e183–e191. 6. US Food and Drug Administration. Guidance for industry—population pharmacokinetics, 1999. Available at: <http://www.fda.gov/downloads/drugs/guidances/UCM072137.pdf>. Accessed on: 01 March 2023. 7. European Medicines Agency. Guideline on reporting the results of population pharmacokinetic analyses, 2007. Available at: http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2009/09/WC500003067.pdf. Accessed on: 01 March 2023. 8. Admiraal R, van Kesteren C, Jol-van der Zijde CM, *et al.* Association between anti-thymocyte globulin exposure and CD4+ immune reconstitution in paediatric haemopoietic cell transplantation: A multicentre, retrospective pharmacodynamic cohort analysis. *Lancet Haematol*. 2015;2(5):e194–e203. 9. Admiraal R, Nierkens S, Bierings MB, *et al.* Individualised dosing of anti-thymocyte globulin in paediatric unrelated allogeneic haematopoietic stem-cell transplantation (PARACHUTE): A single-arm, phase 2 clinical trial. *Lancet Haematol*. 2022;9(2):e111–e120. 10. Drozdov D, Long SL, Gupta AO, *et al.* Bodyweight and absolute lymphocyte count based dosing of rabbit anti-thymocyte globulin in early CD4+ immune reconstitution in patients with inborn errors of metabolism undergoing umbilical cord blood transplantation. *Blood*. 2022;140(1):4797–4798. 11. Srinivasan A, Shah R, Anderson M, *et al.* Retrospective review of use of individualized dosing of rabbit anti-thymocyte globulin on outcomes in pediatric post allogeneic stem cell transplant patients: A single center experience. *Biol Blood Marrow Transplant*. 2019;25(3):S196–S197. 12. Wang H, Zhao Y, Fang S, *et al.* Optimal active anti-thymocyte globulin exposure associated with minimum risk of virus reactivation and comparable acute graft-versus-host disease under adult myeloablative haploidentical peripheral blood stem cell transplantation. *Transplant Cell Ther*. 2022;28(6):332.e1–332.e10.

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